

Whole-Genome Sequences of 26 *Vibrio cholerae* Isolates

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The human pathogen *Vibrio cholerae* employs several adaptive mechanisms for environmental persistence, including natural transformation and type VI secretion, creating a reservoir for the spread of disease. Here, we report whole-genome sequences of 26 diverse *V. cholerae* isolates, significantly increasing the sequence diversity of publicly available *V. cholerae* genomes.

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Vibrio cholerae is a Gram-negative, facultative anaerobe commonly found in salt water and can cause the fatal diarrheal disease cholera. *V. cholerae* is spread predominantly by fecal contamination of water and food sources in both endemic and epidemic regions (1). Governmental and nongovernmental labs worldwide collect *V. cholerae* isolates for standard bacterial surveillance, and many clinical isolates of *V. cholerae* have already been sequenced (2–4). Clinical samples are often isolated from

limited geographic regions and clonally derived with few or no genetic differences (5–8). An expanded characterization of genomes from environmental isolates of *V. cholerae*, which tend to be far more genetically and phenotypically diverse (9), should substantially increase the available sequence diversity of this important human pathogen.

Vibrio spp. are known to encode type VI secretion systems (T6SS), which are often described as bacterial weapons designed

TABLE 1 List of *V. cholerae* strains sequenced in this study

| Strain ^a | Location | Source | Yr of isolation | Type VI killing activity ^b | NCBI accession no. |
|---------------------|----------------------------|--------------|-----------------|---------------------------------------|--------------------|
| 1496-86 | United States (LA) | Moore swab | 1986 | – | MIPC00000000 |
| 2523-87 | United States (LA) | Moore swab | 1974 | + | MIPB00000000 |
| VC48 | United States (FL) | Oyster | 1981 | + | MIOU00000000 |
| 2633-78 | Brazil | Sewage | 1978 | + | MIPH00000000 |
| 857 | Bangladesh | Water | 1996 | + | MIKH00000000 |
| 3272-78 | United States (MD) | Water | 1977 | + | MIOZ00000000 |
| TP | United States (CA) | Water | 2000 | + | MIPK00000000 |
| 2559-78 | United States (LA) | Crab | 1978 | + | MIOU00000000 |
| HE46 | Haiti (center) | Gray water | 2011 | + | MIPM00000000 |
| 2479-86 | United States (LA) | Moore swab | 1986 | + | MIPB00000000 |
| 2497-86 | United States (LA) | Moore swab | 1987 | + | MIPD00000000 |
| 2512-86 | United States (LA) | Moore swab | 1986 | + | MIOY00000000 |
| 2631-78 | United States (LA) | Moore swab | 1978 | + | MIOX00000000 |
| VC22 | United States (FL) | Oyster | 1981 | + | MIKK00000000 |
| VC53 | United States (AL) | Oyster | 2009 | + | MIOU00000000 |
| VC56 | United States (AL) | Oyster | 2009 | + | MIOV00000000 |
| 3568-07 | Mexico | Queso fresco | 2007 | + | MIPL00000000 |
| 1074-78 | Brazil | Sewage | 1978 | + | MIPG00000000 |
| 3223-74 | Guam | Storm drain | 1974 | + | MIZG00000000 |
| 3225-74 | Guam | Storm drain | 1974 | + | MIPF00000000 |
| 2740-80 | United States (Gulf Coast) | Water | 1980 | + | MIKI00000000 |
| 692-79 | United States (LA) | Water | 1979 | + | MIPA00000000 |
| SIO | United States (CA) | Water | 2000 | + | MIPJ00000000 |
| C6706 | Peru | Patient | 1991 | – | MIPJ00000000 |
| MZO-2 | Bangladesh | Patient | 2001 | – | MIKJ00000000 |
| V52 | Sudan | Patient | 1968 | + | MIPN00000000 |

^a Strains were isolated from an environmental source, except strains C6706, MZO-2, and V52.

^b Presence (+) or absence (–) of constitutive type VI killing activity.

to pierce the membranes of adjacent cells and deliver toxic effectors that can lead to lysis of target (prey) cells. In a recent survey, Bernardy et al. (10) noted key differences within a diverse set of isolates for several phenotypes, including chitinase production, contact-dependent killing indicative of T6SS activity, and natural transformation, which can promote horizontal gene transfer. Both clinical and environmental isolates were rarely naturally transformable. In contrast, the majority of environmental, but not clinical, isolates constitutively killed *Escherichia coli* prey. Because different regulatory schemes control the phenotypes tested (11, 12), we sought to better understand the genetics that underlie these diverse *V. cholerae* phenotypes by characterizing whole-genome sequences of 23 environmental and three clinical isolates from Bernardy et al.

All strains were grown overnight in LB medium (Difco) at 37°C, with shaking. Genomic DNA was isolated using a ZR fungal/bacterial DNA mini prep kit (Zymo Research), and paired-end fragment libraries were constructed using a Nextera XT DNA library preparation kit (Illumina) with a fragment length of 300 bp. Libraries were sequenced by the High Throughput Sequencing Core at Georgia Institute of Technology on an Illumina HiSeq 2500 Rapid platform, producing approximately 280 million 100-bp reads in total. Reads were trimmed using Trimmomatic (13) to remove adapters and bases with a read quality score of <20. Genomes were assembled using SPAdes version 3.5 (14) and annotated using the Rapid Annotation and Subsystem Technology (RAST) web tool provided by the National Microbial Pathogen Data Resource (15–18). T6SS genes were annotated using T6SS Predictor (A. T. Chande et al., unpublished data).

T6SS loci were annotated in all genomes in an effort to characterize the genetic basis of T6SS-mediated killing among diverse environmental *V. cholerae* isolates. All genomes were found to encode the previously characterized large cluster and two auxiliary clusters, which together comprise the canonical T6SS loci. In addition, two previously unreported T6SS loci were discovered in six of the isolates. Numerous examples of novel effector-immunity protein pairs, which function together to catalyze T6SS-mediated killing, were characterized among the set of environmental isolate genomes. Taken together, our genome analysis illuminates the diverse repertoire of genetic mechanisms that underlie T6SS-mediated killing in *V. cholerae*.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession numbers listed in Table 1.

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